Supplementary Materials

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Structure-based prediction of HDAC6 substrates validated by enzymatic assay reveals determinants of promiscuity and detects new potential substrates

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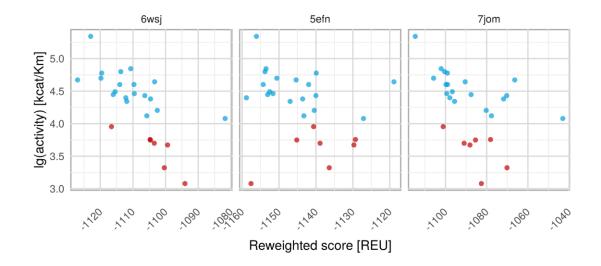
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Supplementary Table S2. Kinetic parameters of the H3 K14_{Ac} peptide and full-

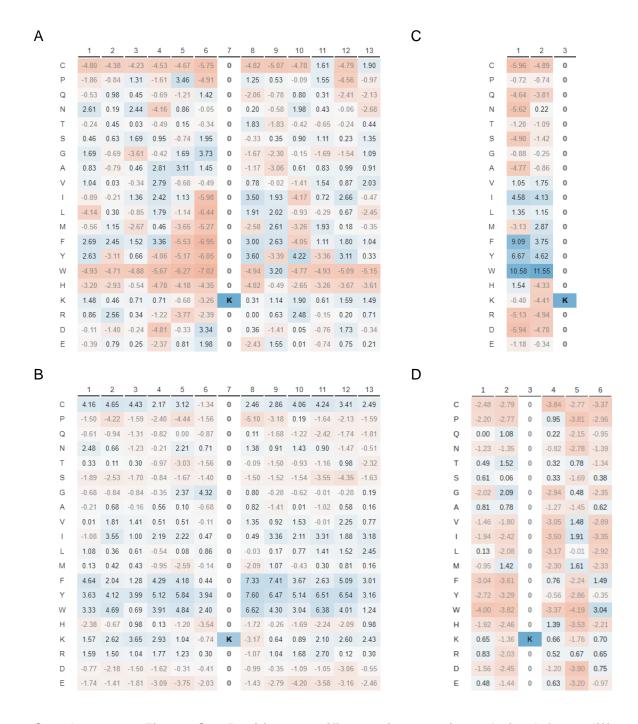
length protein from which it was derived. Related to Table 1.

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Supplementary Figure S1. Performance on different starting structures

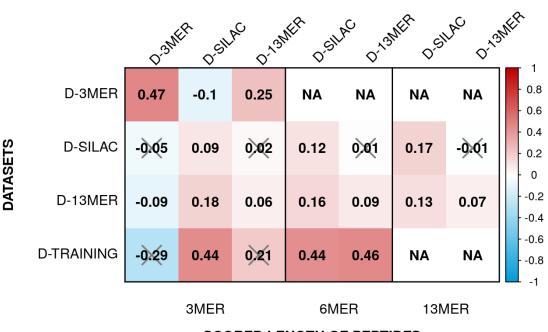
Shown are results for the training set (D-TRAINING). The calibrated protocol was run on a structure of HDAC6 DD2 domain bound to (1) a cyclic peptide (6WSJ; same as **Figure 2A**, (2) a trimer peptide connected to coumarine (5EFN), and (3) a small molecule inhibitor (7JOM). (blue: substrates; red: non-substrates).



Supplementary Figure S2. Position specific scoring matrices derived from different datasets

PSSMS were generated from **A)** D-SILAC, **B)** D-13MER, **C)** D-3MER, and **D)** D-TRAINING sets by applying PSSMSearch to the substrate list of each experiment (for definition of substrates, see **Supplementary Table S1**). (blue: enriched residues, red: depleted amino acids).

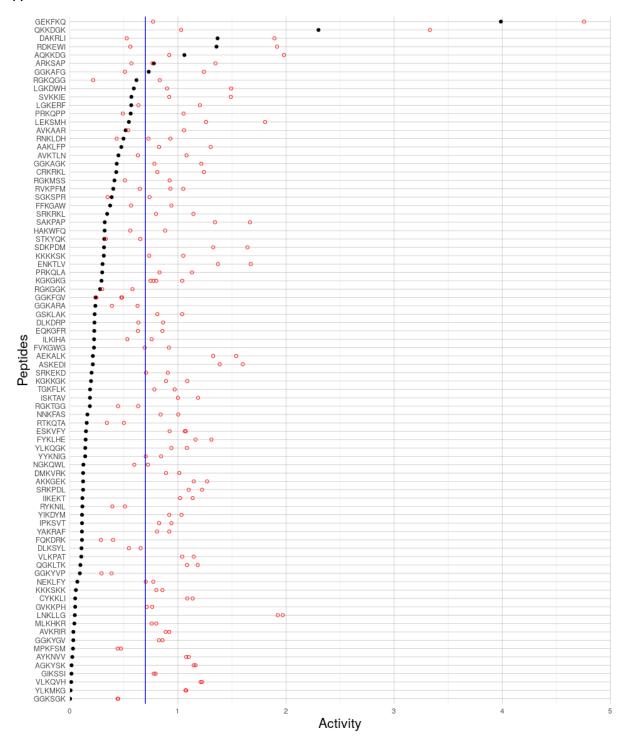
PSSM USED FOR SCORING

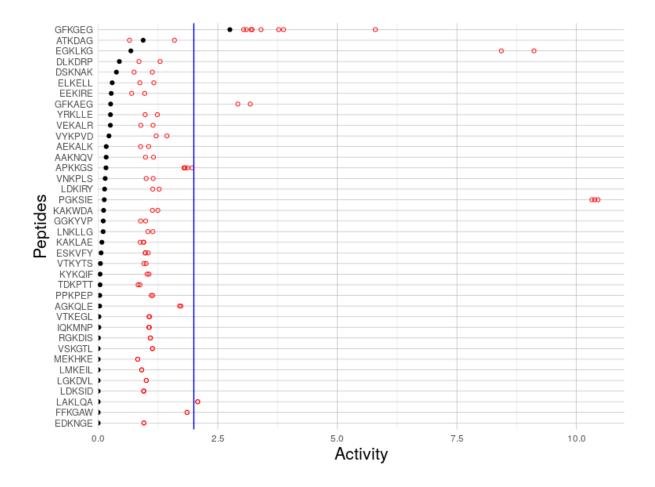


SCORED LENGTH OF PEPTIDES

Supplementary Figure S3. PSSM scores and experimental values do not correlate.

Each dataset was cross-scored with every obtained PSSM depicted in **Supplementary Figure S2**. Three different lengths for scoring (3-, 6- and 13-mers) were used where applicable. D-3MER, D-SILAC and D-13MER labels at the top represent the dataset that the respective PSSM was derived from. 3MER, 6MER and 13MER labels indicate the number of amino acids used for scoring. X indicates non-significant correlations.





Supplementary Figure S4. Range of measured activities for peptides sharing identical core hexamers with differing flanking regions.

A) D-13MER and **B)** D-SILAC datasets. (Black dots: range, red circles: measured values, blue line: threshold of substrate-non-substrate distinction) Related to **Figure 4**.

Supplementary Table S1. Datasets evaluated in this study. The unit of measurement of substrate activity and parameters used to define substrates are indicated. See Text for more details. Related to **Figure 4**.

Dataset reference	N^b peptides	N _{substrates}	N _{non-substrates}	Cutoff for substrates	Cutoff for non-substrates	Unit
D-TRAINING ^a	26	19	7	≧10 ⁴	-	k₀at/Kм
D-CAPPED ^a	16	16	0	≧10 ⁴	-	k₀at/Kм
D-13MER ¹	6797	395 (51) ^c	4127 (245)°	4 out of 4	1 out of 4	# experiments that identified a substrate
D-HPLC ¹	24	17	7	≧10 ⁴	-	$k_{\text{cat}}/K_{\text{M}}$
D-SILAC ²	929	63 (18) ^c	328 (278) ^c	≧2	≦1	ratio (H/L)
D-3MER ³	361	35	33	≧70	≦21	- (intensity)

^a D-TRAINING & D-CAPPED: present study (see **Table 1**).

^b N indicates the number of respective peptides

 $^{^{\}rm c}$ In parentheses: peptides that were reported both in the D-13MER and D-SILAC sets.

Supplementary Table S2. Kinetic parameters of the H3 K14_{Ac} peptide and full-length protein from which it was derived. Related to Table 1.

H3 K14ac substrate	k _{cat} /K _M (M ⁻¹ s ⁻¹)	k _{cat} (s ⁻¹)	<i>K</i> _M (μM)
13-mer: RKSTGG(K-ac)APRKQL	140,000±10,000	4.1 ± 0.3	28 ± 5
Full length protein	80,000±40,000	1.4 ± 0.6	20 ± 10

Supplementary Table S3. Peptides measured for HDAC6 deacetylation in this study but not used for training or validation.

These peptides were not included due to poor measurement accuracy, or peptide length beyond 6 residues. *calculated from only 3 data points. Related to **Table 1**.

Peptide	Protein (site of modification)	<i>k</i> _{cat} / <i>K</i> _M (M⁻¹s⁻¹)	k cat (s ⁻¹)	<i>Κ</i> _M (μM)	Rosetta reweighted score [REU]
ME (K-Ac) KKE	GBP7 (K-389)	3,300 ± 200*	>0.3*	>150*	-1,105
QD(K-Ac)PLR	CCDC86 (K-261)	>2,000*	0.11 ± 0.04*	<50*	-1,092
kgGA(K-Ac)RHR	H4K16 (K-16) ⁴	70,000 ± 20,000	1.23 ± 0.09	17 ± 6	-1,106
SLG(K-Ac)DWHR	CRIP1 (K-22), CRIP1 (K-144)	31,000 ± 5,000	10 ± 2	300 ± 200	-1,113
rMF(K-Ac)QFNK	TRIM28 (K-770)	21,000 ± 1,000	>2	>200	-1,110
riIL(K-Ac)ASR	MSH2 (K-635) ⁵	20,000 ± 3,000	4 ± 1	180 ± 80	-1,115

Supplementary Table S4. Performance of protocol on high-throughput datasets.

(MCC: Matthews correlation coefficient, AUC: area under the ROC curve)

Dataset	Specificity	Sensitivity	MCC	Spearman correlation	AUC
D-13MER	0.18	0.77	-0.05	-0.01	0.49
D-SILAC	0.19	0.89	0.05	-0.05	0.52

Supplementary Table S5. Files of Rosetta protocols.

Constraint File for docking and minimization measured on PDB ID 6WSJ. (Chain A: receptor, chain F is the peptide). Fixbb resfile is used for peptide threading.

Constraint file	Dihedral N 3F CA 3F C 3F N 4F CIRCULARHARMONIC -36.7 0.5 AtomPair OG 531A N 3F HARMONIC 2.9 0.2 AtomPair OD2 705A OH 3F HARMONIC 3.7 0.2 AtomPair OD1 705A OH 3F HARMONIC 5.2 0.2 AtomPair OD2 612A OH 3F HARMONIC 3.4 0.2 AtomPair OD1 612A OH 3F HARMONIC 3.7 0.2 AtomPair ND1 614A OH 3F HARMONIC 3.8 0.2 AtomPair ND1 614A OH 3F HARMONIC 3.8 0.2 AtomPair O 582A NZ 3F HARMONIC 3.3 0.2 AtomPair CE1 583A CD 3F HARMONIC 4.0 0.2 AtomPair CE2 583A CD 3F HARMONIC 3.7 0.2 AtomPair CD2 643A CG 3F HARMONIC 4.0 0.2 AtomPair CD1 643A CG 3F HARMONIC 4.0 0.2 AtomPair CD1 643A CG 3F HARMONIC 3.7 0.2
fixxb resfile	NATRO start 1 F PIKAA E EX 1 EX 2 USE_INPUT_SC 2 F PIKAA G EX 1 EX 2 USE_INPUT_SC 4 F PIKAA F EX 1 EX 2 USE_INPUT_SC 5 F PIKAA V EX 1 EX 2 USE_INPUT_SC 6 F PIKAA R EX 1 EX 2 USE_INPUT_SC

Supplementary Table S6. Commands and flags of Rosetta runs.

All commands were run with Rosetta v2020.28.

Prepack	\$ROSETTA_HOME/main/source/bin/FlexPepDocking.default.linuxgccrelease - s input/6wsj.pdb -ex1 -ex2aro -use_input_sc -flexpep_prepack -nstruct 1 -scorefile ppk.score.sc -flexpep_score_only -out:path:pdb input - out:path:score output -unboundrot input/5eem.pdb 6wsj.pdb
Peptide docking	\$ROSETTA_HOME/main/source/bin/FlexPepDocking.mpi.linuxgccrelease -s input/6wsj.ppk.pdb -ex1 -ex2aro -use_input_sc -constraints:cst_fa_file input/constraints_6wsj.cst -constraints:cst_fa_weight 1.0 -unboundrot input/5eem.pdb input/6wsj.ppk.pdb -nstruct 250 -flexpep_score_only - scorefile refine.score.sc -out:path:pdb output -out:file:silent output/decoys.silent -out:file:silent_struct_type binary - out:path:score output -overwrite -flexPepDocking:pep_refine - lowres_preoptimize (-min_receptor_bb flag was added in some protocols, as described in Methods)
Peptide threading	\$ROSETTA_HOME/main/source/bin/fixbb.default.linuxgccrelease -database \$ROSETTA_HOME/database -resfile resfile -s template.pdb -ex1 -ex2aro - use_input_sc -scorefile design.score.sc -nstruct 1 -unboundrot 5eem.pdb template.pdb
Minimization	\$ROSETTA_HOME/main/source/bin/FlexPepDocking.mpi.linuxgccrelease -s start.ppk.pdb -ex1 -ex2 -ex3 -ex4 -constraints:cst_fa_file constraints.cst -constraints:cst_fa_weight 1.0 -scorefile min.score.sc -unboundrot 5eem.pdb start.ppk.pdb -flexPepDockingMinimizeOnly - flexpep_score_only (-min_receptor_bb flag was added in some protocols, as described in Methods)

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